Item #: SE1-EK-96

BioFront MonoTrace™ Sesame ELISA Kit

Store contents at 2-8˚C

A Monoclonal Antibody-Based Enzyme-Linked Immunosorbent Assay (ELISA) for the Quantitative and/or Qualitative Detection of Sesame in Food

Read instructions carefully before using kit

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DESCRIPTION AND INTENDED USE

The BioFront Technologies Sesame ELISA Kit is an enzyme-linked immunosorbent assay (ELISA) for the quantitative or qualitative detection of sesame. This monoclonal antibody (MAb)-based assay provides a highly sensitive and specific method for the quantification of sesame presence/contamination within a variety of food matrices. It may also be used for simple “yes-no” qualitative assessments. The kit enables a facile and quantitative measure of sesame without the cross-reactivity issues often associated with polyclonal antibody (PAb)-based ELISA kits. The target indicator protein is a major sesame allergen and was selected based on its strong resistance to food processing and high abundance in the sesame seed.

SPECIFICATIONS

***Testing Time (post-extraction):**	**~30 minutes**
Number of Test Wells per Kit:	**96**
****Limit of Detection:**	**0.3 ppm**
*****Range of Quantification:**	**1 to 40 ppm**
Specificity:	**Sesame**
Cross-Reactivity	At concentrations as high as 100,000 ppm, **no cross-reactivity** was observed in a large panel of assayed samples, including: **walnut, almond, coconut, pistachio, pecan, soy, macadamia nut, Brazil nut, and pine nut** ***~10% cross-reactivity observed with hazelnut***
Recovery	Recovery of spiked sesame samples at various concentrations was efficient (>90%) when the following food matrices were assayed: **chocolate, cereals, cake, cookies, ice cream**

Note- calculations of parameters are based on representative data from multiple assays using 10-minute incubation steps.

*Longer sample and conjugate incubation times may increase assay sensitivity, however attention needs to be given to increased background signal that may also result.

**The LOD was determined statistically based on the standard deviation of the response (SD) and assay background according to the formula: LOD = background + 3X SD.

***The range of quantitation (ROQ) was determined experimentally, whereby the lower limit of the ROQ is defined as the lowest concentration at which the assay can reliably and accurately quantify almond in a sample. For quantitation above 40 ppm, samples should be diluted such that the results fall within the ROQ (1 to 40 ppm).

REQUIRED MATERIALS

Kit contents

Reagent	Amount	Use
Assay plate, one 96-well plate	Twelve 8-well strips	Sufficient to obtain 96 assay values
10X ExB = 10X extraction buffer	50 mL	Sufficient to extract > 96 100 mg samples
5X SD = 5X sample diluent	25 mL	Sufficient to dilute samples and standards for > 96 assays
Ready-to-use sesame standards	1 ml each of 40; 20; 10; 4; 1 and 0 ppm	Sufficient for up to five standard curves
10X WB = 10X wash buffer	25 mL	Sufficient wash buffer for > 96 wells
CON = 1X anti-sesame HRP-conjugate	12 mL	Sufficient conjugate for > 96 wells
SUB = high sensitivity TMB substrate	12 mL	Sufficient for > 96 assays
STOP = HRP quench solution	12 mL	Sufficient for > 96 assays

Recommended equipment

- Chamber or waterbath for 60°C incubation
- Balance or scale capable of measuring milligram quantities
- Single and multichannel pipettes, capable of measuring 1-1,000 µL
- Pipet-Aid (or equivalent) and serological pipettes, capable of measuring 5-50 mL
- Reagent reservoirs
- 1.5 or 2.0 mL microfuge tubes and tube racks
- 15 or 50 mL conical tubes
- Distilled water or equivalent
- Absorbent paper or ELISA plate washer
- Tube rotator
- Centrifuge, capable of 2,000 x g
- Vortex
- Microplate reader, capable of reading absorbance at 450 nm

SESAME ASSAY PROCEDURE

It is important for the user to read all instructions carefully before performing the assay. Reagents, including the assay plate, should be equilibrated to room temperature for 30 minutes prior to use.

Important notes

Extraction

The sesame ELISA assay is extremely sensitive, capable of detecting minute amounts of sesame. Careful consideration should be given for the preparation of food matrices to ensure several important parameters:

1. The equipment used to prepare samples must be thoroughly cleaned to prevent the contamination of subsequent samples.
2. Disposable plasticware (tubes, pipette tips, etc.) are used wherever possible.
3. The samples are homogenized completely to prevent excessive intra-sample variation.
4. The supplied extraction buffer is sufficient for up to fifty 1-gram or 1-milliliter samples. If other quantities are used, a 10:1 buffer/sample ratio should be maintained (9 volumes of extraction buffer to 1 part sample).
5. Extractions can be performed at room temperature with good efficiency for most matrices. To achieve maximum assay sensitivity, a heated extraction is recommended.
6. Food matrices containing high concentrations of solid fats, such as chocolate, may require additional heating to completely melt the sample before adding preheated extraction buffer.
7. Food matrices such as spices, dark chocolate or those containing Tannins require the addition of 5% non-fat dry milk to the extraction buffer in order to achieve optimal results. If performance issues with a certain matrix are suspected, please contact a BioFront Technologies representative.

Spike control preparation (optional)

Some food matrices may alter the recovery and sensitivity of the ELISA. If suspected, unspiked and sesame-spiked control matrices can be tested. Values obtained on test samples can then be adjusted accordingly.

Preparation of reagents (extraction buffer, sample diluent, and wash buffer)

Determine the amounts of reagents needed. Dilute the concentrated extraction buffer (**10X ExB**), sample diluent (**5X SD**), and wash buffer **(10X WB)** using distilled water or equivalent. Preheat diluted extraction buffer to 60°C (140°F) prior to use. Maintain diluted sample diluent and wash buffer at room temperature.

Preparation of samples

To insure adequate sampling, it is important that the samples be thoroughly homogenized and the particle size rendered as small as possible. Blending/grinding to a fine powder/flour is strongly recommended. Small particle size also enhances extraction efficiency.

Solid/Liquid sample extraction (read 'important notes' section prior to this step)

1. Transfer 1 gram of finely ground food matrix or 1 ml of liquid sample to a ≥ 15 ml tube.
2. Add 9 ml of preheated 1X extraction buffer (diluted **ExB)** to mixture and briefly vortex to suspend the contents. If other starting quantities are used, a 10:1 buffer/sample ratio should be maintained.
3. Incubate tubes at 60°C (140°F) for 10 minutes, mixing vigorously every ~2 minutes.
4. Spin extraction samples at 2,000 x g for 10 minutes at room temperature and transfer the aqueous phase into a fresh tube for testing.

Swab sample extraction

1. Collection of environmental swab samples from surfaces should be carried out according to the manufacturer's protocol. If using the BioFront Technologies Swab Sample Kit, simply follow the provided sample collection instructions and proceed with the protocol below.
2. Add 4 ml of preheated 1X extraction buffer (diluted **ExB)** to the swab sample collection tube containing the 1 ml swab sample solution and detached swab head. Briefly vortex to mix the contents
3. Incubate tubes at 60°C (140°F) for 10 minutes, mixing vigorously every ~2 minutes.
4. The sample is now ready for testing using the recommended procedure below.

Recommended ELISA procedure

1. Determine the number of assay wells needed for test samples and for standards. Carefully remove the strips that are **not** to be used by gently pushing them from beneath the plate until they pop out and return to Mylar bag. Seal and store at 2-8°C.
2. Dilute sample extracts 1:2 in 1X sample diluent (diluted **SD**).
3. Add 200 µL of diluted samples and ready-to-use standards to the appropriate wells.
4. Incubate plate at room temperature for 10 minutes.
5. Discard well contents, blot onto absorbent paper with a slapping action (or autowash). Wash 3X with 1X wash buffer, (diluted **WB**) using ≥ 200 µL per wash, and blot dry.
6. Add 100 µL of 1X anti- sesame antibody-conjugate (**CON**) to each well.

7. Place plate in dark environment and incubate at room temperature for 10 minutes.

8. Discard well contents, wash, and blot dry as described in step #5.

9. Add 100 µL of HRP substrate (**SUB**) per well.

10. Incubate plate in dark for 10 minutes.

11. Add 100 µL of quench solution (**STOP**) to each well and mix by gently pipetting so as to prevent bubbles that could interfere with absorbance readings.

12. Read the absorbance of the wells using a plate reader programmed with a primary absorbance filter of 450 nm and a differential filter of 630 nm. For some plate readers, the differential filter may be automatically accounted for and reading only at 450 nm will be required. Please consult your reader user manual for more information.

13. Plot the standard curve. Interpolate unknown data using the standard curve and appropriate dilution factor. If the recommended ELISA procedure above is followed, the resulting dilution factor would be two (2). Background may be subtracted to normalize data if desired.

ANALYSIS OF RESULTS

Qualitative analysis of assay results

A qualitative assessment can be made using one or more of the provided assay standards. Any of these standards can be used to define a specific threshold at which the unknown sample can be compared. Samples with normalized absorbance values above the threshold are determined to be positive, whereas those samples below the threshold are determined to be negative.

For swab samples, quantification of the target is generally not performed as the assay, is instead, intended to provide a qualitative indication as to whether or not sesame is present or absent in the test sample. The BioFront Technologies Surface Swabbing Kit has been validated to recover at least 1 microgram sesame protein from a 25 cm^2 area using the swab method.

Quantitative analysis of assay results

A **standard curve** should be generated from the averaged ODs of the 0-40 ppm standards after subtracting the 0 ppm averaged background values. A third-order polynomial (cubic) curve fit is recommended for this evaluation. The ppm concentration of test samples can be determined by plotting OD values onto the curve and multiplying the calculated concentration by the appropriate dilution factor (if used). Note that the ppm designations on the provide standards are intended to allow the direct calculated ppm of total sesame seed in an original food sample.

Performance indications

The supplied sesame standards should yield OD values in line with those indicated on the accompanying lot specific certificate of analysis. Significant deterioration in signal or increased background readings may indicate expiration of the reagents. If quantification is required and the OD of the test sample is above that of the 40 ppm sesame standard, further dilution of the sample should be performed prior to repeating the assay to ensure results fall within the assays ROQ.

Assay claims

When performed as instructed, the assay is capable of a simple yes/no qualitative assessment of sesame presence in food samples or a quantitative determination of sesame content. Extracted food samples that generate a colorimetric readout can be compared to the linear portion of a standard curve, allowing the interpolation of sesame content in ppm. The assay is capable of quantifying sesame content between 1 and 40 ppm.

A negative result by this or any other immunological assay does not assure the complete absence of sesame within the sample. The sample may contain sesame below the limit of detection of this kit. The sesame ELISA kit **does not claim** that food is safe for consumption based upon a determination of sesame content.

SHELF LIFE

Each plate is packed in a vacuum-sealed Mylar™ pouch with desiccant and oxygen absorbing packets to extend the shelf life of the product to a minimum of six (6) months from the date of manufacture, if stored at 4°C. The stability of the ready-to-use standards may deteriorate over time, as indicated in the certificate of analysis accompanying each kit. It is therefore recommended that standards be run and a standard curve generated soon after receiving the kit. For purposes of accurate quantification, the standard curve generated from the user's initial analysis can be used for future experiments. It is, however, recommended that at least one positive control (i.e. 40 ppm sesame standard) and the negative control (0 ppm sesame standard) be run during each analysis to ensure that assay procedure was properly followed and no user errors were committed. BioFront Technologies can also provide fresh standards at the customer's request. The performance of the plates can be adversely affected by excessive exposure to light, moisture, and air. It is recommended that the foil pouch and contents be brought to room temperature before removing the contents to avoid condensation.

MSDS INFORMATION

Material safety data sheets are available on the BioFront website, www.biofronttech.com.

WARRANTY

These products are warranted to perform as described herein. All returns must be pre-approved for refund or credit by a BioFront technical representative and are subject to inspection and verification of contents. Failure to comply may result in a delayed or voided refund.

CUSTOMER SERVICE

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